

Package ‘tabula’

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Title Analysis and Visualization of Archaeological Count Data

Version 3.0.0

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Description An easy way to examine archaeological count data. This package provides several tests and measures of diversity: heterogeneity and evenness (Brillouin, Shannon, Simpson, etc.), richness and rarefaction (Chao1, Chao2, ACE, ICE, etc.), turnover and similarity (Brainerd-Robinson, etc.). It allows to easily visualize count data and statistical thresholds: rank vs abundance plots, heatmaps, Ford (1962) and Bertin (1977) diagrams, etc.

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URL <https://packages.tesselle.org/tabula/>,
<https://github.com/tesselle/tabula>

BugReports <https://github.com/tesselle/tabula/issues>

Depends R (>= 3.5)

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'index_diversity.R' 'index_heterogeneity.R'
'index_rarefaction.R' 'index_richness.R' 'index_similarity.R'
'index_test.R' 'index_turnover.R' 'matrigraph.R' 'mutators.R'
'plot_bertin.R' 'plot_diceleraas.R' 'plot_diversity.R'
'plot_ford.R' 'plot_heatmap.R' 'plot_matrix.R' 'plot_rank.R'
'plot_rarefaction.R' 'plot_spot.R' 'reexport.R' 'seriograph.R'
'show.R' 'statistics.R' 'subset.R' 'tabula-deprecated.R'
'tabula-internal.R' 'tabula-package.R' 'validate.R' 'zzz.R'

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aves	<i>Birds Species and Abundances</i>
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Description

A dataset of birds species and abundances in managed and unmanaged areas along the River Wye (UK).

Usage

aves

Format

A `data.frame` with 2 rows and 26 variables (bird species).

Source

Magurran, A. E. (1988). *Ecological Diversity and its Measurement*. Princeton, NJ: Princeton University Press. doi:10.1007/9789401573580.

See Also

Other datasets: [cantabria](#), [pueblo](#), [woodland](#)

bootstrap_diversity	<i>Bootstrap Estimation</i>
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Description

Samples randomly from the elements of object with replacement.

Usage

```
## S4 method for signature 'DiversityIndex'  
bootstrap(object, n = 1000, f = NULL)
```

Arguments

object	An R object (typically a DiversityIndex object).
n	A non-negative integer giving the number of bootstrap replications.
f	A function that takes a single numeric vector (the result of <code>do</code>) as argument.

Value

If `f` is `NULL` (the default), `bootstrap()` returns a named numeric vector with the following elements:

`original` The observed value of `do` applied to object.

`mean` The bootstrap estimate of mean of `do`.

`bias` The bootstrap estimate of bias of `do`.

`error` The bootstrap estimate of standard error of `do`.

If `f` is a function, `bootstrap()` returns the result of `f` applied to the `n` values of `do`.

Author(s)

N. Frerebeau

See Also

Other resampling methods: [jackknife_diversity](#), [resample\(\)](#)

Examples

```
## Data from Conkey 1980, Kintigh 1989
data("cantabria")

## Shannon diversity index
(h <- heterogeneity(cantabria, method = "shannon"))

## Bootstrap resampling
bootstrap(h, f = NULL)

bootstrap(h, f = summary)

quant <- function(x) quantile(x, probs = c(0.25, 0.50))
bootstrap(h, f = quant)

## Jackknife resampling
jackknife(h)

bootstrap(h, f = summary)
```

cantabria

Early Magdalenian Engraved Bones

Description

A dataset of design elements in engraved bones from Cantabrian Spain.

Usage

cantabria

Format

A [data.frame](#) with 5 rows and 44 variables (designs).

Source

Conkey, M. W. (1980). The Identification of prehistoric hunter-gatherer aggregation sites: The case of Altamira. *Current Anthropology*, 21(5), 609-630.

Kintigh, K. W. (1989). Sample Size, Significance, and Measures of Diversity. In Leonard, R. D. and Jones, G. T., *Quantifying Diversity in Archaeology*. New Directions in Archaeology. Cambridge: Cambridge University Press, p. 25-36.

See Also

Other datasets: [aves](#), [pueblo](#), [woodland](#)

heterogeneity

Heterogeneity and Evenness

Description

- `heterogeneity()` returns an heterogeneity or dominance index.
- `evenness()` returns an evenness measure.

Usage

```
heterogeneity(object, ...)
```

```
evenness(object, ...)
```

```
index_berger(x, ...)
```

```
index_boone(x, ...)
```

```
index_brillouin(x, ...)
```

```
index_mcintosh(x, ...)
```

```
index_shannon(x, ...)
```

```
index_simpson(x, ...)
```

```
## S4 method for signature 'matrix'
```

```

heterogeneity(
  object,
  method = c("berger", "boone", "brillouin", "mcintosh", "shannon", "simpson"),
  j = NULL
)

## S4 method for signature 'data.frame'
heterogeneity(
  object,
  method = c("berger", "boone", "brillouin", "mcintosh", "shannon", "simpson"),
  j = NULL
)

## S4 method for signature 'matrix'
evenness(object, method = c("shannon", "brillouin", "mcintosh", "simpson"))

## S4 method for signature 'data.frame'
evenness(object, method = c("shannon", "brillouin", "mcintosh", "simpson"))

## S4 method for signature 'numeric'
index_berger(x, na.rm = FALSE, ...)

## S4 method for signature 'matrix'
index_boone(x, j = NULL, na.rm = FALSE, ...)

## S4 method for signature 'numeric'
index_brillouin(x, evenness = FALSE, na.rm = FALSE, ...)

## S4 method for signature 'numeric'
index_mcintosh(x, evenness = FALSE, na.rm = FALSE, ...)

## S4 method for signature 'numeric'
index_shannon(x, evenness = FALSE, base = exp(1), na.rm = FALSE, ...)

## S4 method for signature 'numeric'
index_simpson(x, evenness = FALSE, na.rm = FALSE, ...)

```

Arguments

object	A $m \times p$ numeric matrix or data.frame of count data (absolute frequencies giving the number of individuals for each category, i.e. a contingency table). A data.frame will be coerced to a numeric matrix via data.matrix() .
...	Currently not used.
x	A numeric vector of count data (absolute frequencies).
method	A character string specifying the index to be computed (see details). Any unambiguous substring can be given.
j	An integer giving the index of the reference type/taxa. If NULL (the default), the most frequent type/taxa in any assemblage will be used.

na.rm	A numeric scalar: should missing values (including NaN) be removed?
evenness	A logical scalar: should an evenness measure be computed instead of an heterogeneity/dominance index?
base	A positive numeric value specifying the base with respect to which logarithms are computed.

Details

Diversity measurement assumes that all individuals in a specific taxa are equivalent and that all types are equally different from each other (Peet 1974). A measure of diversity can be achieved by using indices built on the relative abundance of taxa. These indices (sometimes referred to as non-parametric indices) benefit from not making assumptions about the underlying distribution of taxa abundance: they only take relative abundances of the species that are present and species richness into account. Peet (1974) refers to them as indices of *heterogeneity*.

Diversity indices focus on one aspect of the taxa abundance and emphasize either *richness* (weighting towards uncommon taxa) or *dominance* (weighting towards abundant taxa; Magurran 1988).

Evenness is a measure of how evenly individuals are distributed across the sample.

Value

- `heterogeneity()` returns an [HeterogeneityIndex](#) object.
- `evenness()` returns an [EvennessIndex](#) object.
- `index_*` return a [numeric](#) vector.

Heterogeneity and Evenness Measures

The following heterogeneity index and corresponding evenness measures are available (see Magurran 1988 for details):

`berger` Berger-Parker dominance index. The Berger-Parker index expresses the proportional importance of the most abundant type. This metric is highly biased by sample size and richness, moreover it does not make use of all the information available from sample.

`boone` Boone heterogeneity measure.

`brillouin` Brillouin diversity index. The Brillouin index describes a known collection: it does not assume random sampling in an infinite population. Pielou (1975) and Laxton (1978) argues for the use of the Brillouin index in all circumstances, especially in preference to the Shannon index.

`mcintosh` McIntosh dominance index. The McIntosh index expresses the heterogeneity of a sample in geometric terms. It describes the sample as a point of a S -dimensional hypervolume and uses the Euclidean distance of this point from the origin.

`shannon` Shannon-Wiener diversity index. The Shannon index assumes that individuals are randomly sampled from an infinite population and that all taxa are represented in the sample (it does not reflect the sample size). The main source of error arises from the failure to include all taxa in the sample: this error increases as the proportion of species discovered in the sample declines (Peet 1974, Magurran 1988). The maximum likelihood estimator (MLE) is used for the relative abundance, this is known to be negatively biased by sample size.

simpson Simpson dominance index for finite sample. The Simpson index expresses the probability that two individuals randomly picked from a finite sample belong to two different types. It can be interpreted as the weighted mean of the proportional abundances. This metric is a true probability value, it ranges from 0 (perfectly uneven) to 1 (perfectly even).

The berger, mcintosh and simpson methods return a *dominance* index, not the reciprocal or inverse form usually adopted, so that an increase in the value of the index accompanies a decrease in diversity.

Note

Ramanujan approximation is used for $x!$ computation if $x > 170$.

Author(s)

N. Frerebeau

References

- Berger, W. H. & Parker, F. L. (1970). Diversity of Planktonic Foraminifera in Deep-Sea Sediments. *Science*, 168(3937), 1345-1347. doi:10.1126/science.168.3937.1345.
- Boone, J. L. (1987). Defining and Measuring Midden Catchment. *American Antiquity*, 52(2), 336-45. doi:10.2307/281785.
- Brillouin, L. (1956). *Science and information theory*. New York: Academic Press.
- Kintigh, K. W. (1989). Sample Size, Significance, and Measures of Diversity. In Leonard, R. D. and Jones, G. T., *Quantifying Diversity in Archaeology*. New Directions in Archaeology. Cambridge: Cambridge University Press, p. 25-36.
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- McIntosh, R. P. (1967). An Index of Diversity and the Relation of Certain Concepts to Diversity. *Ecology*, 48(3), 392-404. doi:10.2307/1932674.
- Peet, R. K. (1974). The Measurement of Species Diversity. *Annual Review of Ecology and Systematics*, 5(1), 285-307. doi:10.1146/annurev.es.05.110174.001441.
- Pielou, E. C. (1975). *Ecological Diversity*. New York: Wiley. doi:10.4319/lo.1977.22.1.0174b
- Shannon, C. E. (1948). A Mathematical Theory of Communication. *The Bell System Technical Journal*, 27, 379-423. doi:10.1002/j.15387305.1948.tb01338.x.
- Simpson, E. H. (1949). Measurement of Diversity. *Nature*, 163(4148), 688-688. doi:10.1038/163688a0.

See Also

Other diversity measures: [occurrence\(\)](#), [rarefaction\(\)](#), [richness\(\)](#), [similarity\(\)](#), [simulate\(\)](#), [turnover\(\)](#)

Examples

```
## Data from Conkey 1980, Kintigh 1989
data("cantabria")

## Shannon diversity index
(h <- heterogeneity(cantabria, method = "shannon"))
(e <- evenness(cantabria, method = "shannon"))

plot(h)
```

jackknife_diversity *Jackknife Estimation*

Description

Jackknife Estimation

Usage

```
## S4 method for signature 'DiversityIndex'
jackknife(object, f = NULL)
```

Arguments

object An R object (typically a [DiversityIndex](#) object).

f A [function](#) that takes a single numeric vector (the leave-one-out values of do) as argument.

Value

If **f** is NULL (the default), `jackknife()` returns a named numeric vector with the following elements:

original The observed value of do applied to object.

mean The jackknife estimate of mean of do.

bias The jackknife estimate of bias of do.

error The jackknife estimate of standard error of do.

If **f** is a function, `jackknife()` returns the result of **f** applied to the leave-one-out values of do.

Author(s)

N. Frerebeau

See Also

Other resampling methods: [bootstrap_diversity](#), [resample\(\)](#)

Examples

```
## Data from Conkey 1980, Kintigh 1989
data("cantabria")

## Shannon diversity index
(h <- heterogeneity(cantabria, method = "shannon"))

## Bootstrap resampling
bootstrap(h, f = NULL)

bootstrap(h, f = summary)

quant <- function(x) quantile(x, probs = c(0.25, 0.50))
bootstrap(h, f = quant)

## Jackknife resampling
jackknife(h)

bootstrap(h, f = summary)
```

matrigraph

Matrigraph

Description

- `matrigraph()` produces a heatmap highlighting the deviations from independence.
- `pvi()` computes for each cell of a numeric matrix the percentage to the column theoretical independence value.

Usage

```
matrigraph(object, ...)
```

```
pvi(object, ...)
```

```
## S4 method for signature 'matrix'
pvi(object)
```

```
## S4 method for signature 'data.frame'
pvi(object)
```

```
## S4 method for signature 'matrix'
matrigraph(object, reverse = FALSE, axes = TRUE, ...)
```

```
## S4 method for signature 'data.frame'
matrigraph(object, reverse = FALSE, ...)
```

Arguments

object	A $m \times p$ numeric <code>matrix</code> or <code>data.frame</code> of count data (absolute frequencies giving the number of individuals for each category, i.e. a contingency table).
...	Currently not used.
reverse	A <code>logical</code> scalar: should negative deviations be centered (see details)?
axes	A <code>logical</code> scalar: should axes be drawn on the plot?

Details

PVI (in french "pourcentages de valeur d'indépendance") is calculated for each cell as the percentage to the column theoretical independence value: PVI greater than 1 represent positive deviations from the independence, whereas PVI smaller than 1 represent negative deviations (Desachy 2004).

The PVI matrix allows to explore deviations from independence (an intuitive approach to χ^2), in such a way that a high-contrast matrix has quite significant deviations, with a low risk of being due to randomness (Desachy 2004).

`matrigraph()` displays the deviations from independence:

- If the PVI is equal to 1 (statistical independence), the cell of the matrix is filled in grey.
- If the PVI is less than 1 (negative deviation from independence), the size of the grey square is proportional to the PVI (the white margin thus represents the fraction of negative deviation).
- If the PVI is greater than 1 (positive deviation), a black square representing the fraction of positive deviations is superimposed. For large positive deviations (PVI greater than 2), the cell is filled in black.

If `reverse` is `TRUE`, the fraction of negative deviations is displayed as a white square.

Value

- `matrigraph()` is called it for its side-effects: it results in a graphic being displayed (invisibly returns object).
- `pvi()` returns a `numeric matrix`.

Author(s)

N. Frerebeau

References

Desachy, B. (2004). Le sériographe EPPM: un outil informatisé de sériation graphique pour tableaux de comptages. *Revue archéologique de Picardie*, 3(1), 39-56. doi:10.3406/pica.2004.2396.

See Also

`plot_heatmap()`

Other plot methods: `plot_bertin()`, `plot_diceleraas()`, `plot_diversity`, `plot_ford()`, `plot_heatmap()`, `plot_rank()`, `plot_rarefaction`, `plot_spot()`, `seriograph()`

Examples

```
## Data from Desachy 2004
data("compiegne", package = "folio")

## Matrigraph
matrigraph(compiegne)
matrigraph(compiegne, reverse = TRUE)

## Compute PVI
counts_pvi <- pvi(compiegne)
plot_heatmap(counts_pvi, col = khroma::color("iridescent")(12))
```

mutators

Get or Set Parts of an Object

Description

Getters and setters to extract or replace parts of an object.

Usage

```
get_method(x)

## S4 method for signature 'DiversityIndex'
labels(object)

## S4 method for signature 'RarefactionIndex'
labels(object)

## S4 method for signature 'DiversityIndex'
get_method(x)
```

Arguments

object, x An R object from which to get or set element(s).

Value

- `labels()` returns a suitable set of labels from an object for use in printing or plotting.

Author(s)

N. Frerebeau

occurrence	<i>Co-Occurrence</i>
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Description

Co-Occurrence

Usage

```
occurrence(object, ...)  
  
## S4 method for signature 'matrix'  
occurrence(object)  
  
## S4 method for signature 'data.frame'  
occurrence(object)
```

Arguments

object	A $m \times p$ numeric matrix or data.frame of count data (absolute frequencies giving the number of individuals for each category, i.e. a contingency table). A data.frame will be coerced to a numeric matrix via data.matrix() .
...	Currently not used.

Details

A co-occurrence matrix is a symmetric matrix with zeros on its main diagonal, which works out how many times each pairs of taxa/types occur together in at least one sample.

Value

A [stats::dist](#) object.

Author(s)

N. Frerebeau

See Also

Other diversity measures: [heterogeneity\(\)](#), [rarefaction\(\)](#), [richness\(\)](#), [similarity\(\)](#), [simulate\(\)](#), [turnover\(\)](#)

Examples

```
## Data from Conkey 1980, Kintigh 1989
data("cantabria")

## Plot spot diagram of a co-occurrence matrix
occ <- occurrence(cantabria)
plot_spot(occ)
```

plot_bertin

Bertin Diagram

Description

Plots a Bertin diagram.

Usage

```
plot_bertin(object, ...)
```

S4 method for signature 'matrix'

```
plot_bertin(
  object,
  threshold = NULL,
  freq = FALSE,
  margin = 1,
  col = c("white", "black"),
  flip = TRUE,
  axes = TRUE,
  ...
)
```

S4 method for signature 'data.frame'

```
plot_bertin(
  object,
  threshold = NULL,
  freq = FALSE,
  margin = 1,
  col = c("white", "black"),
  flip = TRUE,
  axes = TRUE,
  ...
)
```

Arguments

object A $m \times p$ numeric [matrix](#) or [data.frame](#) of count data (absolute frequencies giving the number of individuals for each category, i.e. a contingency table).

...	Currently not used.
threshold	A function that takes a numeric vector as argument and returns a numeric threshold value (see below). If NULL (the default), no threshold is computed. Only used if freq is FALSE.
freq	A logical scalar indicating whether conditional proportions given margins should be used (i.e. entries of object, divided by the appropriate marginal sums).
margin	An integer vector giving the margins to split by: 1 indicates individuals/rows (the default), 2 indicates variables/columns. Only used if freq is TRUE.
col	A vector of colors.
flip	A logical scalar: should x and y axis be flipped? Defaults to TRUE.
axes	A logical scalar: should axes be drawn on the plot?

Value

plot_bertin() is called it for its side-effects: it results in a graphic being displayed (invisibly returns object).

Bertin Matrix

As de Falguerolles *et al.* (1997) points out: "In abstract terms, a Bertin matrix is a matrix of displays. ... To fix ideas, think of a data matrix, variable by case, with real valued variables. For each variable, draw a bar chart of variable value by case. High-light all bars representing a value above some sample threshold for that variable."

Author(s)

N. Frerebeau

References

Bertin, J. (1977). *La graphique et le traitement graphique de l'information*. Paris: Flammarion. Nouvelle Bibliothèque Scientifique.

de Falguerolles, A., Friedrich, F. & Sawitzki, G. (1997). A Tribute to J. Bertin's Graphical Data Analysis. In W. Badilla & F. Faulbaum (eds.), *SoftStat '97: Advances in Statistical Software 6*. Stuttgart: Lucius & Lucius, p. 11-20.

See Also

Other plot methods: [matrigraph\(\)](#), [plot_diceleraas\(\)](#), [plot_diversity](#), [plot_ford\(\)](#), [plot_heatmap\(\)](#), [plot_rank\(\)](#), [plot_rarefaction](#), [plot_spot\(\)](#), [seriograph\(\)](#)

Examples

```
## Data from Lipo et al. 2015
data("mississippi", package = "folio")

## Plot a Bertin diagram...
```

```
## ...without threshold
plot_bertin(mississippi)

## ...with the variable mean as threshold
plot_bertin(mississippi, threshold = mean)

## Plot conditional proportions
plot_bertin(mississippi, freq = TRUE, margin = 1)
plot_bertin(mississippi, freq = TRUE, margin = 2)
```

plot_diceleraas	<i>Dice-Leraas Diagram</i>
-----------------	----------------------------

Description

Plots a Dice-Leraas diagram.

Usage

```
plot_diceleraas(object, ...)

## S4 method for signature 'matrix'
plot_diceleraas(
  object,
  main = NULL,
  sub = NULL,
  ann = graphics::par("ann"),
  axes = TRUE,
  frame.plot = FALSE,
  panel.first = NULL,
  panel.last = NULL,
  ...
)

## S4 method for signature 'data.frame'
plot_diceleraas(
  object,
  main = NULL,
  sub = NULL,
  ann = graphics::par("ann"),
  axes = TRUE,
  frame.plot = FALSE,
  panel.first = NULL,
  panel.last = NULL,
  ...
)
```


Arguments

object	A $m \times p$ numeric <code>matrix</code> or <code>data.frame</code> of count data (absolute frequencies giving the number of individuals for each category, i.e. a contingency table). A <code>data.frame</code> will be coerced to a numeric matrix via <code>data.matrix()</code> .
...	Further <code>graphical parameters</code> .
main	A <code>character</code> string giving a main title for the plot.
sub	A <code>character</code> string giving a subtitle for the plot.
ann	A <code>logical</code> scalar: should the default annotation (title and x, y and z axis labels) appear on the plot?
axes	A <code>logical</code> scalar: should axes be drawn on the plot?
frame.plot	A <code>logical</code> scalar: should a box be drawn around the plot?
panel.first	An expression to be evaluated after the plot axes are set up but before any plotting takes place. This can be useful for drawing background grids.
panel.last	An expression to be evaluated after plotting has taken place but before the axes, title and box are added.

Details

In a Dice-Leraas diagram, the horizontal line represents the range of data (min-max) and the small vertical line indicates the mean. The black rectangle is twice the standard error on the mean, while the white rectangle is one standard deviation on either side of the mean.

Value

`plot_diceleraas()` is called it for its side-effects: it results in a graphic being displayed (invisibly returns object).

Author(s)

N. Frerebeau

References

- Dice, L. R., & Leraas, H. J. (1936). A Graphic Method for Comparing Several Sets of Measurements. *Contributions from the Laboratory of Vertebrate Genetics*, 3: 1-3.
- Hubbs, C. L., & C. Hubbs (1953). An Improved Graphical Analysis and Comparison of Series of Samples. *Systematic Biology*, 2(2): 49-56. doi:10.2307/sysbio/2.2.49.
- Simpson, G. G., Roe, A., & Lewontin, R. C. *Quantitative Zoology*. New York: Harcourt, Brace and Company, 1960.

See Also

Other plot methods: `matrigraph()`, `plot_bertin()`, `plot_diversity`, `plot_ford()`, `plot_heatmap()`, `plot_rank()`, `plot_rarefaction`, `plot_spot()`, `seriograph()`

Examples

```
## Data from Desachy 2004
data("compiegne", package = "folio")

## Plot a Dice-Leraas diagram
plot_diceleraas(compiegne)
```

plot_diversity	<i>Diversity Plot</i>
----------------	-----------------------

Description

Diversity Plot

Usage

```
## S4 method for signature 'DiversityIndex,missing'
plot(
  x,
  log = "x",
  col.mean = "#DDAA33",
  col.interval = "#004488",
  lty.mean = "solid",
  lty.interval = "dashed",
  lwd.mean = 1,
  lwd.interval = 1,
  main = NULL,
  sub = NULL,
  ann = graphics::par("ann"),
  axes = TRUE,
  frame.plot = axes,
  panel.first = NULL,
  panel.last = NULL,
  ...
)
```

Arguments

x A [DiversityIndex](#) object to be plotted.

log A [character](#) string indicating which axes should be in log scale. Defaults to x.

col.mean, col.interval A [character](#) string specifying the color of the lines.

lty.mean, lty.interval A [character](#) string or [numeric](#) value specifying the line types.

lwd.mean, lwd.interval A non-negative [numeric](#) value specifying the line widths.

main	A character string giving a main title for the plot.
sub	A character string giving a subtitle for the plot.
ann	A logical scalar: should the default annotation (title and x, y and z axis labels) appear on the plot?
axes	A logical scalar: should axes be drawn on the plot?
frame.plot	A logical scalar: should a box be drawn around the plot?
panel.first	An an expression to be evaluated after the plot axes are set up but before any plotting takes place. This can be useful for drawing background grids.
panel.last	An expression to be evaluated after plotting has taken place but before the axes, title and box are added.
...	Further graphical parameters to be passed to graphics::points() , particularly, cex, col and pch.

Value

plot() is called it for its side-effects: it results in a graphic being displayed (invisibly returns x).

Author(s)

N. Frerebeau

See Also

Other plot methods: [matrigraph\(\)](#), [plot_bertin\(\)](#), [plot_diceleraas\(\)](#), [plot_ford\(\)](#), [plot_heatmap\(\)](#), [plot_rank\(\)](#), [plot_rarefaction](#), [plot_spot\(\)](#), [seriograph\(\)](#)

Examples

```
## Data from Conkey 1980, Kintigh 1989
data("cantabria")

## Assemblage diversity size comparison
## Warning: this may take a few seconds!
h <- heterogeneity(cantabria, method = "shannon")
h_sim <- simulate(h)
plot(h_sim)

r <- richness(cantabria, method = "count")
r_sim <- simulate(r)
plot(r_sim)
```

`plot_ford`*Ford Diagram*

Description

Plots a Ford (battleship curve) diagram.

Usage

```
plot_ford(object, ...)  
  
## S4 method for signature 'matrix'  
plot_ford(  
  object,  
  weights = FALSE,  
  EPPM = FALSE,  
  fill = "darkgrey",  
  border = NA,  
  axes = TRUE,  
  ...  
)  
  
## S4 method for signature 'data.frame'  
plot_ford(object, weights = FALSE, EPPM = FALSE)
```

Arguments

<code>object</code>	A $m \times p$ numeric <code>matrix</code> or <code>data.frame</code> of count data (absolute frequencies giving the number of individuals for each category, i.e. a contingency table).
<code>...</code>	Currently not used.
<code>weights</code>	A <code>logical</code> scalar: should the row sums be displayed?
<code>EPPM</code>	A <code>logical</code> scalar: should the EPPM be drawn? See <code>seriograph()</code> .
<code>fill</code>	The color for filling the bars.
<code>border</code>	The color to draw the borders.
<code>axes</code>	A <code>logical</code> scalar: should axes be drawn on the plot?

Value

`plot_ford()` is called it for its side-effects: it results in a graphic being displayed (invisibly returns object).

Author(s)

N. Frerebeau

References

Ford, J. A. (1962). *A quantitative method for deriving cultural chronology*. Washington, DC: Pan American Union. Technical manual 1.

See Also

Other plot methods: [matrigraph\(\)](#), [plot_bertin\(\)](#), [plot_diceleraas\(\)](#), [plot_diversity](#), [plot_heatmap\(\)](#), [plot_rank\(\)](#), [plot_rarefaction](#), [plot_spot\(\)](#), [seriograph\(\)](#)

Examples

```
## Data from Lipo et al. 2015
data("mississippi", package = "folio")

## Plot a Ford diagram
plot_ford(mississippi)

plot_ford(mississippi, weights = TRUE)
```

plot_heatmap	<i>Heatmap</i>
--------------	----------------

Description

Plots a heatmap.

Usage

```
plot_heatmap(object, ...)

## S4 method for signature 'matrix'
plot_heatmap(
  object,
  col = grDevices::hcl.colors(12, "YlOrBr", rev = TRUE),
  diag = TRUE,
  upper = TRUE,
  lower = TRUE,
  freq = FALSE,
  margin = 1,
  axes = TRUE,
  legend = TRUE,
  ...
)

## S4 method for signature 'data.frame'
plot_heatmap(
  object,
```

```

col = grDevices::hcl.colors(12, "YlOrBr", rev = TRUE),
diag = TRUE,
upper = TRUE,
lower = TRUE,
freq = FALSE,
margin = 1,
axes = TRUE,
legend = TRUE,
...
)

## S4 method for signature 'dist'
plot_heatmap(
  object,
  col = grDevices::hcl.colors(12, "YlOrBr", rev = TRUE),
  diag = FALSE,
  upper = FALSE,
  lower = !upper,
  axes = TRUE,
  legend = TRUE,
  ...
)

```

Arguments

object	A $m \times p$ numeric matrix or data.frame of count data (absolute frequencies giving the number of individuals for each category, i.e. a contingency table).
...	Currently not used.
col	A vector of colors.
diag	A logical scalar indicating whether the diagonal of the matrix should be plotted. Only used if object is a symmetric matrix.
upper	A logical scalar indicating whether the upper triangle of the matrix should be plotted. Only used if object is a symmetric matrix.
lower	A logical scalar indicating whether the lower triangle of the matrix should be plotted. Only used if object is a symmetric matrix.
freq	A logical scalar indicating whether conditional proportions given margins should be used (i.e. entries of object, divided by the appropriate marginal sums).
margin	An integer vector giving the margins to split by: 1 indicates individuals/rows (the default), 2 indicates variables/columns. Only used if freq is TRUE.
axes	A logical scalar: should axes be drawn on the plot?
legend	A logical scalar: should a legend be displayed?

Value

plot_heatmap() is called it for its side-effects: it results in a graphic being displayed (invisibly returns object).

Author(s)

N. Frerebeau

See Also

Other plot methods: [matrigraph\(\)](#), [plot_bertin\(\)](#), [plot_diceleraas\(\)](#), [plot_diversity](#), [plot_ford\(\)](#), [plot_rank\(\)](#), [plot_rarefaction](#), [plot_spot\(\)](#), [seriograph\(\)](#)

Examples

```
## Data from Conkey 1980, Kintigh 1989
data("cantabria")

## Plot raw data
plot_heatmap(cantabria)

## Plot conditional proportions
plot_heatmap(cantabria, freq = TRUE, margin = 1)
plot_heatmap(cantabria, freq = TRUE, margin = 2)
```

plot_rank

Rank Plot

Description

Plots a rank vs relative abundance diagram.

Usage

```
plot_rank(object, ...)
```

S4 method for signature 'matrix'

```
plot_rank(
  object,
  log = NULL,
  main = NULL,
  sub = NULL,
  ann = graphics::par("ann"),
  axes = TRUE,
  frame.plot = axes,
  panel.first = NULL,
  panel.last = NULL,
  legend = list(x = "topright"),
  ...
)
```

S4 method for signature 'data.frame'

```
plot_rank(
```

```

object,
log = NULL,
main = NULL,
sub = NULL,
ann = graphics::par("ann"),
axes = TRUE,
frame.plot = axes,
panel.first = NULL,
panel.last = NULL,
legend = list(x = "topright"),
...
)

```

Arguments

object	A $m \times p$ numeric matrix or data.frame of count data (absolute frequencies giving the number of individuals for each category, i.e. a contingency table). A data.frame will be coerced to a numeric matrix via data.matrix() .
...	Further graphical parameters .
log	A character string which contains "x" if the x axis is to be logarithmic, "y" if the y axis is to be logarithmic and "xy" or "yx" if both axes are to be logarithmic (base 10).
main	A character string giving a main title for the plot.
sub	A character string giving a subtitle for the plot.
ann	A logical scalar: should the default annotation (title and x, y and z axis labels) appear on the plot?
axes	A logical scalar: should axes be drawn on the plot?
frame.plot	A logical scalar: should a box be drawn around the plot?
panel.first	An an expression to be evaluated after the plot axes are set up but before any plotting takes place. This can be useful for drawing background grids.
panel.last	An expression to be evaluated after plotting has taken place but before the axes, title and box are added.
legend	A list of additional arguments to be passed to graphics::legend() ; names of the list are used as argument names. If NULL, no legend is displayed.

Value

plot_rank() is called it for its side-effects: it results in a graphic being displayed (invisibly returns object).

Author(s)

N. Frerebeau

References

Magurran, A. E. (1988). *Ecological Diversity and its Measurement*. Princeton, NJ: Princeton University Press. [doi:10.1007/9789401573580](https://doi.org/10.1007/9789401573580).

See Also

Other plot methods: [matrigraph\(\)](#), [plot_bertin\(\)](#), [plot_diceleraas\(\)](#), [plot_diversity](#), [plot_ford\(\)](#), [plot_heatmap\(\)](#), [plot_rarefaction](#), [plot_spot\(\)](#), [seriograph\(\)](#)

Examples

```
## Data from Conkey 1980, Kintigh 1989
data("cantabria")

## Plot rank vs abundance
plot_rank(cantabria)

## Change graphical parameters
col <- khroma::color("bright")(5)
plot_rank(cantabria, col = col, pch = 15:19, lty = 2)
```

plot_rarefaction	<i>Rarefaction Plot</i>
------------------	-------------------------

Description

Rarefaction Plot

Usage

```
## S4 method for signature 'RarefactionIndex,missing'
plot(
  x,
  main = NULL,
  sub = NULL,
  ann = graphics::par("ann"),
  axes = TRUE,
  frame.plot = axes,
  panel.first = NULL,
  panel.last = NULL,
  legend = list(x = "topleft"),
  ...
)
```

Arguments

x	A RarefactionIndex object to be plotted.
main	A character string giving a main title for the plot.
sub	A character string giving a subtitle for the plot.
ann	A logical scalar: should the default annotation (title and x, y and z axis labels) appear on the plot?

axes	A logical scalar: should axes be drawn on the plot?
frame.plot	A logical scalar: should a box be drawn around the plot?
panel.first	An an expression to be evaluated after the plot axes are set up but before any plotting takes place. This can be useful for drawing background grids.
panel.last	An expression to be evaluated after plotting has taken place but before the axes, title and box are added.
legend	A list of additional arguments to be passed to graphics::legend() ; names of the list are used as argument names. If NULL, no legend is displayed.
...	Further graphical parameters to be passed to graphics::lines() .

Value

plot() is called it for its side-effects: it results in a graphic being displayed (invisibly returns x).

Author(s)

N. Frerebeau

See Also

Other plot methods: [matrigraph\(\)](#), [plot_bertin\(\)](#), [plot_diceleeraas\(\)](#), [plot_diversity](#), [plot_ford\(\)](#), [plot_heatmap\(\)](#), [plot_rank\(\)](#), [plot_spot\(\)](#), [seriograph\(\)](#)

Examples

```
## Data from Conkey 1980, Kintigh 1989
data("cantabria")

## Replicate fig. 3 from Baxter 2011
rare <- rarefaction(cantabria, sample = 23, method = "baxter")
plot(rare, panel.first = graphics::grid())

## Change graphical parameters
col <- khroma::color("bright")(5)
plot(rare, col = col, lty = 1:5)
```

plot_spot

Spot Plot

Description

Plots a spot matrix.

Usage

```
plot_spot(object, ...)  
  
## S4 method for signature 'matrix'  
plot_spot(  
  object,  
  type = c("ring", "plain"),  
  col = grDevices::hcl.colors(12, "YlOrBr", rev = TRUE),  
  diag = TRUE,  
  upper = TRUE,  
  lower = TRUE,  
  freq = FALSE,  
  margin = 1,  
  axes = TRUE,  
  legend = TRUE,  
  ...  
)  
  
## S4 method for signature 'data.frame'  
plot_spot(  
  object,  
  type = c("ring", "plain"),  
  col = grDevices::hcl.colors(12, "YlOrBr", rev = TRUE),  
  diag = TRUE,  
  upper = TRUE,  
  lower = TRUE,  
  freq = FALSE,  
  margin = 1,  
  axes = TRUE,  
  legend = TRUE,  
  ...  
)  
  
## S4 method for signature 'dist'  
plot_spot(  
  object,  
  type = c("ring", "plain"),  
  col = grDevices::hcl.colors(12, "YlOrBr", rev = TRUE),  
  diag = FALSE,  
  upper = FALSE,  
  lower = !upper,  
  axes = TRUE,  
  legend = TRUE,  
  ...  
)
```

Arguments

object	A $m \times p$ numeric matrix or data.frame of count data (absolute frequencies giving the number of individuals for each category, i.e. a contingency table).
...	Currently not used.
type	A character string specifying the graph to be plotted. It must be one of "ring" (the default) or "plain". Any unambiguous substring can be given.
col	A vector of colors.
diag	A logical scalar indicating whether the diagonal of the matrix should be plotted. Only used if object is a symmetric matrix.
upper	A logical scalar indicating whether the upper triangle of the matrix should be plotted. Only used if object is a symmetric matrix.
lower	A logical scalar indicating whether the lower triangle of the matrix should be plotted. Only used if object is a symmetric matrix.
freq	A logical scalar indicating whether conditional proportions given margins should be used (i.e. entries of object, divided by the appropriate marginal sums).
margin	An integer vector giving the margins to split by: 1 indicates individuals/rows (the default), 2 indicates variables/columns. Only used if freq is TRUE.
axes	A logical scalar: should axes be drawn on the plot?
legend	A logical scalar: should a legend be displayed?

Details

The spot matrix can be considered as a variant of the [Bertin diagram](#) where the data are first transformed to relative frequencies.

Value

plot_spot() is called it for its side-effects: it results in a graphic being displayed (invisibly returns object).

Note

Adapted from Dan Gopstein's original [idea](#).

Author(s)

N. Frerebeau

See Also

Other plot methods: [matrigraph\(\)](#), [plot_bertin\(\)](#), [plot_diceleraas\(\)](#), [plot_diversity](#), [plot_ford\(\)](#), [plot_heatmap\(\)](#), [plot_rank\(\)](#), [plot_rarefaction](#), [seriograph\(\)](#)

Examples

```
## Data from Huntley 2004, 2008
data("pueblo")

## Plot spot diagram of count data
plot_spot(pueblo, type = "ring")
plot_spot(pueblo, type = "plain")

## Plot conditional proportions
plot_spot(pueblo, freq = TRUE, margin = 1)
plot_spot(pueblo, freq = TRUE, margin = 2)
```

pueblo	<i>Pueblo IV Period Ceramics</i>
--------	----------------------------------

Description

A dataset of ceramic counts from the Zuni region.

Usage

pueblo

Format

A `data.frame` with 9 rows and 5 variables (compositional groups).

Source

Huntley, D. L. (2004). *Interaction, Boundaries, and Identities: A Multiscalar Approach to the Organizational Scale of Pueblo IV Zuni Society*. Ph.D. Dissertation, Arizona State University.

Huntley, D. L. (2022). *Ancestral Zuni Glaze-Decorated Pottery: Viewing Pueblo IV Regional Organization through Ceramic Production and Exchange*. Anthropological Papers of the University of Arizona 72. Tucson: University of Arizona Press. doi:10.2307/j.ctv2ngx5n8.

See Also

Other datasets: [aves](#), [cantabria](#), [woodland](#)

rarefaction

Rarefaction

Description

Rarefaction

Usage

```
rarefaction(object, ...)
```

```
index_baxter(x, ...)
```

```
index_hurlbert(x, ...)
```

```
## S4 method for signature 'matrix'
```

```
rarefaction(object, sample = NULL, method = c("hurlbert", "baxter"), step = 1)
```

```
## S4 method for signature 'data.frame'
```

```
rarefaction(object, sample = NULL, method = c("hurlbert", "baxter"), step = 1)
```

```
## S4 method for signature 'numeric'
```

```
index_baxter(x, sample, ...)
```

```
## S4 method for signature 'numeric'
```

```
index_hurlbert(x, sample, ...)
```

Arguments

object A $m \times p$ numeric [matrix](#) or [data.frame](#) of count data (absolute frequencies giving the number of individuals for each category, i.e. a contingency table). A [data.frame](#) will be coerced to a numeric matrix via [data.matrix\(\)](#).

... Currently not used.

x A [numeric](#) vector of count data (absolute frequencies).

sample A length-one [numeric](#) vector giving the sub-sample size. The size of sample should be smaller than total community size.

method A [character](#) string or vector of strings specifying the index to be computed (see details). Any unambiguous substring can be given.

step An [integer](#) giving the increment of the sample size.

Value

- `rarefaction()` returns a [RarefactionIndex](#) object.
- `index_*` return a [numeric](#) vector.

Rarefaction Measures

The following rarefaction measures are available for count data:

baxter Baxter's rarefaction.

hurlbert Hurlbert's unbiased estimate of Sander's rarefaction.

Details

The number of different taxa, provides an instantly comprehensible expression of diversity. While the number of taxa within a sample is easy to ascertain, as a term, it makes little sense: some taxa may not have been seen, or there may not be a fixed number of taxa (e.g. in an open system; Peet 1974). As an alternative, *richness* (S) can be used for the concept of taxa number (McIntosh 1967).

It is not always possible to ensure that all sample sizes are equal and the number of different taxa increases with sample size and sampling effort (Magurran 1988). Then, *rarefaction* ($E(S)$) is the number of taxa expected if all samples were of a standard size (i.e. taxa per fixed number of individuals). Rarefaction assumes that imbalances between taxa are due to sampling and not to differences in actual abundances.

Author(s)

N. Frerebeau

References

Baxter, M. J. (2001). Methodological Issues in the Study of Assemblage Diversity. *American Antiquity*, 66(4), 715-725. doi:10.2307/2694184.

Hurlbert, S. H. (1971). The Nonconcept of Species Diversity: A Critique and Alternative Parameters. *Ecology*, 52(4), 577-586. doi:10.2307/1934145.

Sander, H. L. (1968). Marine Benthic Diversity: A Comparative Study. *The American Naturalist*, 102(925), 243-282.

See Also

[plot\(\)](#)

Other diversity measures: [heterogeneity\(\)](#), [occurrence\(\)](#), [richness\(\)](#), [similarity\(\)](#), [simulate\(\)](#), [turnover\(\)](#)

Examples

```
## Data from Conkey 1980, Kintigh 1989
data("cantabria")

## Replicate fig. 3 from Baxter 2011
rare <- rarefaction(cantabria, sample = 23, method = "baxter")
plot(rare, panel.first = graphics::grid())

## Change graphical parameters
col <- khroma::color("bright")(5)
plot(rare, col = col, lty = 1:5)
```

`resample`*Resample*

Description

Simulates observations from a multinomial distribution.

Usage

```
resample(object, ...)
```

```
## S4 method for signature 'numeric'
```

```
resample(object, do, n, size = sum(object), ..., f = NULL)
```

Arguments

<code>object</code>	A numeric vector of count data (absolute frequencies).
<code>...</code>	Extra arguments passed to <code>do</code> .
<code>do</code>	A function that takes <code>object</code> as an argument and returns a single numeric value.
<code>n</code>	A non-negative integer specifying the number of bootstrap replications.
<code>size</code>	A non-negative integer specifying the sample size.
<code>f</code>	A function that takes a single numeric vector (the result of <code>do</code>) as argument.

Value

If `f` is `NULL`, `resample()` returns the `n` values of `do`. Else, returns the result of `f` applied to the `n` values of `do`.

Author(s)

N. Frerebeau

See Also

[stats::rmultinom\(\)](#)

Other resampling methods: [bootstrap_diversity](#), [jackknife_diversity](#)

Examples

```
## Sample observations from a multinomial distribution
x <- sample(1:100, 50, TRUE)
resample(x, do = median, n = 100)
```

```
## Estimate the 25th, 50th and 95th percentiles
quant <- function(x) { quantile(x, probs = c(0.25, 0.50, 0.75)) }
resample(x, n = 100, do = median, f = quant)
```

richness	<i>Richness</i>
----------	-----------------

Description

- richness() returns sample richness.
- composition() returns asymptotic species richness.

Usage

```
richness(object, ...)
```

```
composition(object, ...)
```

```
index_ace(x, ...)
```

```
index_ice(x, ...)
```

```
index_chao1(x, ...)
```

```
index_chao2(x, ...)
```

```
index_margalef(x, ...)
```

```
index_menhinick(x, ...)
```

```
## S4 method for signature 'matrix'
```

```
richness(object, method = c("count", "margalef", "menhinick"))
```

```
## S4 method for signature 'data.frame'
```

```
richness(object, method = c("count", "margalef", "menhinick"))
```

```
## S4 method for signature 'matrix'
```

```
composition(  
  object,  
  method = c("chao1", "ace", "chao2", "ice"),  
  unbiased = FALSE,  
  improved = FALSE,  
  k = 10  
)
```

```
## S4 method for signature 'data.frame'
```

```
composition(  
  object,  
  method = c("chao1", "ace", "chao2", "ice"),  
  unbiased = FALSE,  
  improved = FALSE,
```

```

    k = 10
  )

## S4 method for signature 'numeric'
index_margalef(x, na.rm = FALSE, ...)

## S4 method for signature 'numeric'
index_menhinick(x, na.rm = FALSE, ...)

## S4 method for signature 'numeric'
index_ace(x, k = 10, ...)

## S4 method for signature 'numeric'
index_chao1(x, unbiased = FALSE, improved = FALSE, ...)

## S4 method for signature 'matrix'
index_ice(x, k = 10, ...)

## S4 method for signature 'matrix'
index_chao2(x, unbiased = FALSE, improved = FALSE, ...)

```

Arguments

object	A $m \times p$ numeric matrix or data.frame of count data (absolute frequencies giving the number of individuals for each category, i.e. a contingency table). A data.frame will be coerced to a numeric matrix via data.matrix() .
...	Further arguments to be passed to internal methods.
x	A numeric vector or matrix of count data (absolute frequencies).
method	A character string or vector of strings specifying the index to be computed (see details). Any unambiguous substring can be given.
unbiased	A logical scalar. Should the bias-corrected estimator be used? Only used with "chao1" or "chao2" (improved) estimator.
improved	A logical scalar. Should the improved estimator be used? Only used with "chao1" or "chao2".
k	A length-one numeric vector giving the threshold between rare/infrequent and abundant/frequent species. Only used if method is "ace" or "ice".
na.rm	A numeric scalar: should missing values (including NaN) be removed?

Value

- richness() returns a [RichnessIndex](#) object.
- composition() returns a [CompositionIndex](#) object.
- index_*(*x*) return a [numeric](#) vector.

Details

The number of different taxa, provides an instantly comprehensible expression of diversity. While the number of taxa within a sample is easy to ascertain, as a term, it makes little sense: some taxa may not have been seen, or there may not be a fixed number of taxa (e.g. in an open system; Peet 1974). As an alternative, *richness* (S) can be used for the concept of taxa number (McIntosh 1967).

It is not always possible to ensure that all sample sizes are equal and the number of different taxa increases with sample size and sampling effort (Magurran 1988). Then, *rarefaction* ($E(S)$) is the number of taxa expected if all samples were of a standard size (i.e. taxa per fixed number of individuals). Rarefaction assumes that imbalances between taxa are due to sampling and not to differences in actual abundances.

Richness Measures

The following richness measures are available for count data:

count Returns the number of observed taxa/types.

margalef Margalef richness index.

menhinick Menhinick richness index.

Asymptotic Species Richness

The following measures are available for count data:

ace Abundance-based Coverage Estimator.

chao1 (improved/unbiased) Chao1 estimator.

The following measures are available for replicated incidence data:

ice Incidence-based Coverage Estimator.

chao2 (improved/unbiased) Chao2 estimator.

Author(s)

N. Frerebeau

References

Chao, A. (1984). Nonparametric Estimation of the Number of Classes in a Population. *Scandinavian Journal of Statistics*, 11(4), 265-270.

Chao, A. (1987). Estimating the Population Size for Capture-Recapture Data with Unequal Catchability. *Biometrics* 43(4), 783-791. doi:10.2307/2531532.

Chao, A. & Chiu, C.-H. (2016). Species Richness: Estimation and Comparison. In Balakrishnan, N., Colton, T., Everitt, B., Piegorisch, B., Ruggeri, F. & Teugels, J. L. (Eds.), *Wiley StatRef: Statistics Reference Online*. Chichester, UK: John Wiley & Sons, Ltd., 1-26. doi:10.1002/9781118445112.stat03432.pub2

Chao, A. & Lee, S.-M. (1992). Estimating the Number of Classes via Sample Coverage. *Journal of the American Statistical Association*, 87(417), 210-217. doi:10.1080/01621459.1992.10475194.

Chiu, C.-H., Wang, Y.-T., Walther, B. A. & Chao, A. (2014). An improved nonparametric lower bound of species richness via a modified good-turing frequency formula. *Biometrics*, 70(3), 671-682. doi:10.1111/biom.12200.

Magurran, A. E. (1988). *Ecological Diversity and its Measurement*. Princeton, NJ: Princeton University Press. doi:10.1007/9789401573580.

Kintigh, K. W. (1989). Sample Size, Significance, and Measures of Diversity. In Leonard, R. D. and Jones, G. T., *Quantifying Diversity in Archaeology*. New Directions in Archaeology. Cambridge: Cambridge University Press, p. 25-36.

Magurran, A. E. & Brian J. McGill (2011). *Biological Diversity: Frontiers in Measurement and Assessment*. Oxford: Oxford University Press.

Margalef, R. (1958). Information Theory in Ecology. *General Systems*, 3, 36-71.

Menhinick, E. F. (1964). A Comparison of Some Species-Individuals Diversity Indices Applied to Samples of Field Insects. *Ecology*, 45(4), 859-861. doi:10.2307/1934933.

McIntosh, R. P. (1967). An Index of Diversity and the Relation of Certain Concepts to Diversity. *Ecology*, 48(3), 392-404. doi:10.2307/1932674.

See Also

[plot\(\)](#)

Other diversity measures: [heterogeneity\(\)](#), [occurrence\(\)](#), [rarefaction\(\)](#), [similarity\(\)](#), [simulate\(\)](#), [turnover\(\)](#)

Examples

```
## Data from Magurran 1988, p. 128-129
trap <- matrix(data = c(9, 3, 0, 4, 2, 1, 1, 0, 1, 0, 1, 1,
                      1, 0, 1, 0, 0, 0, 1, 2, 0, 5, 3, 0),
              nrow = 2, byrow = TRUE, dimnames = list(c("A", "B"), NULL))

## Margalef and Menhinick index
richness(trap, method = "margalef") # 2.55 1.88
richness(trap, method = "menhinick") # 1.95 1.66

## Data from Chao & Chiu 2016
brazil <- matrix(
  data = rep(x = c(1:21, 23, 25, 27, 28, 30, 32, 34:37, 41,
                 45, 46, 49, 52, 89, 110, 123, 140),
            times = c(113, 50, 39, 29, 15, 11, 13, 5, 6, 6, 3, 4,
                    3, 5, 2, 5, 2, 2, 2, 2, 1, 2, 1, 1, 1, 1, 1,
                    0, 0, 2, 1, 1, 1, 1, 1, 0, 1, 1, 0, 0)),
  nrow = 1, byrow = TRUE
)

## Chao1-type estimators (asymptotic species richness)
composition(brazil, method = c("chao1"), unbiased = FALSE) # 461.625
composition(brazil, method = c("ace"), k = 10) # 445.822
```

seriograph

Seriograph

Description

- `seriograph()` produces a Ford diagram highlighting the relationships between rows and columns.
- `eppm()` computes for each cell of a numeric matrix the positive difference from the column mean percentage.

Usage

```
seriograph(object, ...)

eppm(object, ...)

## S4 method for signature 'matrix'
eppm(object)

## S4 method for signature 'data.frame'
eppm(object)

## S4 method for signature 'matrix'
seriograph(
  object,
  weights = FALSE,
  fill = "darkgrey",
  border = NA,
  axes = TRUE,
  ...
)

## S4 method for signature 'data.frame'
seriograph(
  object,
  weights = FALSE,
  fill = "darkgrey",
  border = NA,
  axes = TRUE,
  ...
)
```

Arguments

`object` A $m \times p$ numeric `matrix` or `data.frame` of count data (absolute frequencies giving the number of individuals for each category, i.e. a contingency table).

...	Currently not used.
weights	A logical scalar: should the row sums be displayed?
fill	The color for filling the bars.
border	The color to draw the borders.
axes	A logical scalar: should axes be drawn on the plot?

Details

The positive difference from the column mean percentage (in french "écart positif au pourcentage moyen", EPPM) represents a deviation from the situation of statistical independence. As independence can be interpreted as the absence of relationships between types and the chronological order of the assemblages, EPPM is a useful tool to explore significance of relationship between rows and columns related to seriation (Desachy 2004).

`seriograph()` superimposes the frequencies (grey) and EPPM values (black) for each row-column pair in a Ford diagram.

Value

- `seriograph()` is called it for its side-effects: it results in a graphic being displayed (invisibly returns object).
- `eppm()` returns a [numeric matrix](#).

Author(s)

N. Frerebeau

References

Desachy, B. (2004). Le sériographe EPPM: un outil informatisé de sériation graphique pour tableaux de comptages. *Revue archéologique de Picardie*, 3(1), 39-56. doi:10.3406/pica.2004.2396.

See Also

[plot_ford\(\)](#)

Other plot methods: [matrigraph\(\)](#), [plot_bertin\(\)](#), [plot_diceleraas\(\)](#), [plot_diversity](#), [plot_ford\(\)](#), [plot_heatmap\(\)](#), [plot_rank\(\)](#), [plot_rarefaction](#), [plot_spot\(\)](#)

Examples

```
## Data from Desachy 2004
data("compiegne", package = "folio")

## Seriograph
seriograph(compiegne)
seriograph(compiegne, weights = TRUE)

## Compute EPPM
counts_eppm <- eppm(compiegne)
plot_heatmap(counts_eppm, col = khroma::color("Yl0rBr")(12))
```

similarity	<i>Similarity</i>
------------	-------------------

Description

Similarity

Usage

```
similarity(object, ...)  
  
index_jaccard(x, y, ...)  
  
index_sorenson(x, y, ...)  
  
index_bray(x, y, ...)  
  
index_morisita(x, y, ...)  
  
index_brainerd(x, y, ...)  
  
index_binomial(x, y, ...)  
  
## S4 method for signature 'matrix'  
similarity(  
  object,  
  method = c("brainerd", "bray", "jaccard", "morisita", "sorenson", "binomial")  
)  
  
## S4 method for signature 'data.frame'  
similarity(  
  object,  
  method = c("brainerd", "bray", "jaccard", "morisita", "sorenson", "binomial")  
)  
  
## S4 method for signature 'character,character'  
index_jaccard(x, y)  
  
## S4 method for signature 'logical,logical'  
index_jaccard(x, y)  
  
## S4 method for signature 'numeric,numeric'  
index_jaccard(x, y)  
  
## S4 method for signature 'logical,logical'  
index_sorenson(x, y)
```

```
## S4 method for signature 'numeric,numeric'
index_sorenson(x, y)

## S4 method for signature 'numeric,numeric'
index_bray(x, y)

## S4 method for signature 'numeric,numeric'
index_morisita(x, y)

## S4 method for signature 'numeric,numeric'
index_brainerd(x, y)

## S4 method for signature 'numeric,numeric'
index_binomial(x, y)
```

Arguments

object	A $m \times p$ numeric <code>matrix</code> or <code>data.frame</code> of count data (absolute frequencies giving the number of individuals for each category, i.e. a contingency table). A <code>data.frame</code> will be coerced to a numeric matrix via <code>data.matrix()</code> .
...	Currently not used.
x, y	A length- p numeric vector of count data.
method	A <code>character</code> string specifying the method to be used (see details). Any unambiguous substring can be given.

Details

β -diversity can be measured by addressing *similarity* between pairs of samples/cases (Brainerd-Robinson, Jaccard, Morisita-Horn and Sorenson indices). Similarity between pairs of taxa/types can be measured by assessing the degree of co-occurrence (binomial co-occurrence).

Jaccard, Morisita-Horn and Sorenson indices provide a scale of similarity from 0-1 where 1 is perfect similarity and 0 is no similarity. The Brainerd-Robinson index is scaled between 0 and 200. The Binomial co-occurrence assessment approximates a Z-score.

`binomial` Binomial co-occurrence assessment. This assesses the degree of co-occurrence between taxa/types within a dataset. The strongest associations are shown by large positive numbers, the strongest segregations by large negative numbers.

`brainerd` Brainerd-Robinson quantitative index. This is a city-block metric of similarity between pairs of samples/cases.

`bray` Sorenson quantitative index (Bray and Curtis modified version of the Sorenson index).

`jaccard` Jaccard qualitative index.

`morisita` Morisita-Horn quantitative index.

`sorenson` Sorenson qualitative index.

Value

- `similarity()` returns a `stats::dist` object.
- `index_*`() return a `numeric` vector.

Author(s)

N. Frerebeau

References

- Brainerd, G. W. (1951). The Place of Chronological Ordering in Archaeological Analysis. *American Antiquity*, 16(04), 301-313. doi:10.2307/276979.
- Bray, J. R. & Curtis, J. T. (1957). An Ordination of the Upland Forest Communities of Southern Wisconsin. *Ecological Monographs*, 27(4), 325-349. doi:10.2307/1942268.
- Kintigh, K. (2006). Ceramic Dating and Type Associations. In J. Hantman and R. Most (eds.), *Managing Archaeological Data: Essays in Honor of Sylvia W. Gaines*. Anthropological Research Paper, 57. Tempe, AZ: Arizona State University, p. 17-26.
- Magurran, A. E. (1988). *Ecological Diversity and its Measurement*. Princeton, NJ: Princeton University Press. doi:10.1007/9789401573580.
- Robinson, W. S. (1951). A Method for Chronologically Ordering Archaeological Deposits. *American Antiquity*, 16(04), 293-301. doi:10.2307/276978.

See Also

Other diversity measures: [heterogeneity\(\)](#), [occurrence\(\)](#), [rarefaction\(\)](#), [richness\(\)](#), [simulate\(\)](#), [turnover\(\)](#)

Examples

```
## Data from Huntley 2004, 2008
data("pueblo")

## Brainerd-Robinson measure
(C <- similarity(pueblo, "brainerd"))
plot_spot(C)

## Data from Magurran 1988, p. 166
data("aves")

## Jaccard measure (presence/absence data)
similarity(aves, "jaccard") # 0.46

## Sorenson measure (presence/absence data)
similarity(aves, "sorenson") # 0.63

# Jaccard measure (Bray's formula ; count data)
similarity(aves, "bray") # 0.44

# Morisita-Horn measure (count data)
similarity(aves, "morisita") # 0.81
```

`simulate`*Measure Diversity by Comparing to Simulated Assemblages*

Description

Measure Diversity by Comparing to Simulated Assemblages

Usage

```
## S4 method for signature 'DiversityIndex'  
simulate(  
  object,  
  n = 1000,  
  step = 1,  
  interval = c("percentiles", "student", "normal"),  
  level = 0.8,  
  progress = getOption("tabula.progress")  
)
```

Arguments

<code>object</code>	A DiversityIndex object.
<code>n</code>	A non-negative integer giving the number of bootstrap replications.
<code>step</code>	An integer giving the increment of the sample size.
<code>interval</code>	A character string giving the type of confidence interval to be returned. It must be one "percentiles" (sample quantiles, as described in Kintigh 1984; the default), "student" or "normal". Any unambiguous substring can be given.
<code>level</code>	A length-one numeric vector giving the confidence level.
<code>progress</code>	A logical scalar: should a progress bar be displayed?

Value

Returns a [DiversityIndex](#) object.

Author(s)

N. Frerebeau

References

Baxter, M. J. (2001). Methodological Issues in the Study of Assemblage Diversity. *American Antiquity*, 66(4), 715-725. doi:10.2307/2694184.

Kintigh, K. W. (1984). Measuring Archaeological Diversity by Comparison with Simulated Assemblages. *American Antiquity*, 49(1), 44-54. doi:10.2307/280511.

See Also

[plot\(\)](#), [resample\(\)](#)

Other diversity measures: [heterogeneity\(\)](#), [occurrence\(\)](#), [rarefaction\(\)](#), [richness\(\)](#), [similarity\(\)](#), [turnover\(\)](#)

Examples

```
## Data from Conkey 1980, Kintigh 1989
data("cantabria")

## Assemblage diversity size comparison
## Warning: this may take a few seconds!
h <- heterogeneity(cantabria, method = "shannon")
h_sim <- simulate(h)
plot(h_sim)

r <- richness(cantabria, method = "count")
r_sim <- simulate(r)
plot(r_sim)
```

test_diversity

Diversity Test

Description

Compares Shannon diversity between samples.

Usage

```
test_diversity(object, ...)

## S4 method for signature 'matrix'
test_diversity(object, adjust = "holm", ...)

## S4 method for signature 'data.frame'
test_diversity(object, adjust = "holm", ...)
```

Arguments

object	A $m \times p$ numeric matrix or data.frame of count data (absolute frequencies giving the number of individuals for each category, i.e. a contingency table). A data.frame will be coerced to a numeric matrix via data.matrix() .
...	Further arguments to be passed to internal methods.
adjust	A character string specifying the method for adjusting p values (see stats::p.adjust()).

Details

This test produces two sided pairwise comparisons: it returns a matrix of adjusted p values.

Value

A numeric matrix.

Author(s)

N. Frerebeau

References

Magurran, A. E. (1988). *Ecological Diversity and its Measurement*. Princeton, NJ: Princeton University Press. doi:10.1007/9789401573580.

Examples

```
## Data from Conkey 1980, Kintigh 1989
data("cantabria")

## Shannon diversity test
test_diversity(cantabria)
```

turnover

Turnover

Description

Returns the degree of turnover in taxa composition along a gradient or transect.

Usage

```
turnover(object, ...)

index_whittaker(x, ...)

index_cody(x, ...)

index_routledge1(x, ...)

index_routledge2(x, ...)

index_routledge3(x, ...)

index_wilson(x, ...)

## S4 method for signature 'matrix'
```

```

turnover(
  object,
  method = c("whittaker", "cody", "routledge1", "routledge2", "routledge3", "wilson"),
  ...
)

## S4 method for signature 'data.frame'
turnover(
  object,
  method = c("whittaker", "cody", "routledge1", "routledge2", "routledge3", "wilson"),
  ...
)

## S4 method for signature 'matrix'
index_whittaker(x)

## S4 method for signature 'matrix'
index_cody(x)

## S4 method for signature 'matrix'
index_routledge1(x)

## S4 method for signature 'matrix'
index_routledge2(x)

## S4 method for signature 'matrix'
index_routledge3(x)

## S4 method for signature 'matrix'
index_wilson(x)

```

Arguments

object, x	A $m \times p$ numeric matrix or data.frame of count data or incidence data. A data.frame will be coerced to a numeric matrix via data.matrix() .
...	Further arguments to be passed to internal methods.
method	A character string specifying the method to be used (see details). Any unambiguous substring can be given.

Details

The following methods can be used to ascertain the degree of *turnover* in taxa composition along a gradient (β -diversity) on qualitative (presence/absence) data. This assumes that the order of the matrix rows (from 1 to n) follows the progression along the gradient/transect.

whittaker Whittaker measure.

cody Cody measure.

routledge1 Routledge first measure.

routledge2 Routledge second measure.

routledge3 Routledge third measure. This is the exponential form of the second measure.

wilson Wilson measure.

Value

A `numeric` vector.

Author(s)

N. Frerebeau

References

Cody, M. L. (1975). Towards a theory of continental species diversity: Bird distributions over Mediterranean habitat gradients. In M. L. Cody & J. M. Diamond (Eds.), *Ecology and Evolution of Communities*. Cambridge, MA: Harvard University Press, p. 214-257.

Routledge, R. D. (1977). On Whittaker's Components of Diversity. *Ecology*, 58(5), 1120-1127. doi:10.2307/1936932.

Whittaker, R. H. (1960). Vegetation of the Siskiyou Mountains, Oregon and California. *Ecological Monographs*, 30(3), 279-338. doi:10.2307/1943563.

Wilson, M. V., & Shmida, A. (1984). Measuring Beta Diversity with Presence-Absence Data. *The Journal of Ecology*, 72(3), 1055-1064. doi:10.2307/2259551.

See Also

Other diversity measures: `heterogeneity()`, `occurrence()`, `rarefaction()`, `richness()`, `similarity()`, `simulate()`

Examples

```
## Data from Magurran 1988, p. 162
data("woodland")

## Whittaker's measure
turnover(woodland, "whittaker") # 1

## Cody's measure
turnover(woodland, "cody") # 3

## Routledge's measures
turnover(woodland, "routledge1") # 0.29
turnover(woodland, "routledge2") # 0.56
turnover(woodland, "routledge3") # 1.75

## Wilson and Shmida's measure
turnover(woodland, "wilson") # 1
```

woodland

Trees Incidences

Description

A dataset of presence or absence of trees in six (10 x 10 m) quadrats along a transect through a deciduous woodland.

Usage

woodland

Format

A [data.frame](#) with 6 rows (quadrats) and 6 variables (tree species).

Source

Magurran, A. E. (1988). *Ecological Diversity and its Measurement*. Princeton, NJ: Princeton University Press. [doi:10.1007/9789401573580](https://doi.org/10.1007/9789401573580).

See Also

Other datasets: [aves](#), [cantabria](#), [pueblo](#)

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